REMARKS

Applicants hereby submit that the enclosures fulfill the requirements under 37 C.F.R. §1.821-1.825. The amendments in the specification merely insert the paper copy of the Sequence Listing and sequence identifiers in the specification. No new matter has been added.

Attached hereto is a marked-up version of the changes made to the specification by the current amendment.

Please apply any charges or credits to Deposit Account No. 06-1050.

Respectfully submitted,

Date: 1/3/02

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"Version With Markings to Show Changes Made"

In the specification:

Paragraph beginning at page 15, line 4, has been amended as follows:

Figure 2 shows the sequence alignment[21] of the amino acid sequences of MASP-2 (clone phl-4; amino acid residues 16-686 of SEQ ID NO:2), MASP-1[17,22] (SEQ ID NO:6), C1r[23,24] (SEQ ID NO:7) and C1s[25,26] (SEQ ID NO:8).

Paragraph beginning at page 15, line 14, has been amended as follows:

Figure 6 shows the cDNA sequence and deduced amino acid sequence of MASP-2 (SEQ ID NOs:3 and 2, respectively).

Paragraph beginning at page 46, line 10, has been amended as follows:

The liver is the primary site of synthesis of C1r, C1s, and MASP-1. Thus, RNA from liver was used as template for RT-PCR with primers deduced from the obtained peptide sequences. First strand synthesis of cDNA was carried out with 1.3 ìg human liver RNA using a First-Strand cDNA Synthesis Kit (Pharmacia). PCR was performed on this cDNA using degenerate sense and antisense primers derived from the amino acid sequences EYANDQER (SEQ ID NO:4) and KPFTGFEA (SEQ ID NO:5), respectively. The PCR program consisted of 1 cycle with annealing at 50EC; 1 cycle with annealing at 55EC, and 33 cycles with annealing at 60EC. The resulting 300 bp PCR product was cloned into the *E. coli* plasmid pCRII using the TA-cloning kit (InVitrogen) and the nucleotide sequence of the insert was determined.